

ANTI-IVASION EFFECT OF BITTER MELON LEAF EXTRACT ON RAT PROSTATE CANCER CELL LINE

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Introduction: Cancer metastasis remains a major cause of death in cancer patients. Metastasis is a multi-step process. The invasive ability involving the first step of tumor metastasis is a major cause of treatment failure. *Momordica charantia* L. or bitter melon is widely consumed as vegetable and especially folk medicine in Asia. Extracts of bitter melon have been reported to possess anti-tumor activity such as inhibition of mouse spontaneous mammary tumorigenesis and benzo(a)pyrene-induced mice forestomach tumorigenesis, antioxidant activities, antiviral (HIV inhibitor), antidiabetic and immunomodulating properties. Our previous study revealed that bitter melon leaf extract (BMLE) inhibited P-glycoprotein activity and reverse multidrug-resistant in cervical carcinoma cell line (KB-V1), increased the intracellular accumulation of [³H]-vinblastine, decreased the [³H]-vinblastine efflux in KB-V1 cells and increased their sensitivity to vinblastine. However, there is no evidence whether BMLE has effects on prostate cancer cell invasion. **Purpose:** In this study we examined whether BMLE possessed inhibitory effects against invasive of rat prostate cancer cell line (PLS10). **Methods:** To determine cytotoxicity of BMLE on PLS10, WST-1 assay was used. The effect of BMLE on invasiveness and motility of PLS10 was determined by BD Biocoat™ Matrigel™ Invasion Chamber and BD Falcon inserts respectively. Gelatin zymography assay was performed to analyze matrix metalloproteinase (MMP)-2 and MMP-9 secretion from PLS10 cells. MMP-2, MMP-9 and tissue inhibitor of metalloproteinase (TIMP)-2 mRNA levels were analyzed by real time PCR. Collagenase type IV activity was assayed with the EnzChek Gelatinase/Collagenase Assay Kit. The results are presented as means ±SD (% of control) from three independent experiments. **Results:** WST-1 assay showed BMLE decreased cell viability on PLS10 cells (48 h; IC₂₀<75 µg/ml, IC₅₀: <150 µg/ml). 48 hr treatment of BMLE with non-toxic dose (25 and 50 µg/ml) significantly decreased PLS10 cell invasion (51±18; p<0.05 and 42±12; p<0.005 respectively) and gradually decreased cell migration at 24 hr (62±4 and 36±2; p<0.005 respectively). MMP-2 secreted in the culture media was significantly decreased when PLS10 cells were treated with 25 and 50 µg/ml for 24 hr (78±10 and 54±18; p<0.05 respectively) and 50 µg/ml for 48 hr (67±12; p<0.005). After 24 hr 50 µg/ml BMLE treatment, MMP-9 secretion was significantly decreased (73±15; p<0.05). 25 and 50 µg/ml BMLE treatments significantly decreased MMP-9 secretions at 48 hour (83±10; p<0.05 and 71±8; p<0.005 respectively). We further analysis the effect of BMLE on MMP-2, MMP-9 and TIMP-2 gene expression using real time PCR. We found that BMLE (50 µg/ml) significantly decreased gene expression of MMP-2 (24 h; 25±6; p<0.005 and 48 h; 38±27 p<0.005) and MMP-9 (24 h; 39±27; p<0.005 and 48 h; 17±7 p<0.005 respectively) but markedly increase mRNA level of TIMP-2 (10, 25 and 50 µg/ml BMLE treatment; 24 h; 128±12; p<0.05, 153±20; p<0.05, 231±3; p<0.005 and 48 h; 124±8; p<0.05, 128±9; p<0.005, 222±67; p<0.05 respectively) which known to have inhibitory effect on the activity of on MMP-2. Detection of MMP activity was performed by an EnzChek Gelatinase/Collagenase Assay kit, the result showed that collagenase type IV activity was inhibit by BMLE (50 and 100 µg/ml BMLE treatment; 84±1; p<0.005 and 85±5 p<0.05 respectively). **Conclusion:** These results suggest that BMLE exerts anti-invasion effect on PLS10

cells through modulation of MMP-2 and MMP-9 secretion, down-regulated MMP-2 and MMP-9 gene expression, enhanced TIMP-2 gene expression and inhibits collagenase type IV activity. **Acknowledgement:** This work was supported by grants from the Royal Golden Jubilee Ph.D. Program of Thailand and the Society for Promotion of Pathology of Nagoya, Japan.